Original Article

Check for updates

DEFB126 2-nt Deletion (rs11467417) as a Potential Risk Factor for Chlamydia Trachomatis Infection and Subsequent Infertility in Iranian Men

Kaveh Haratian [1], Parnaz Borjian Boroujeni [2], Marjan Sabbaghian [3], Elham Maghareh Abed [2], Maedeh Moazenchi [2], Anahita Mohseni Meybodi [1,2*]

1- Department of Pathology and Laboratory Medicine, Western University, Ontario, Canada
2- Department of Genetics, Reproductive Biomedicine Research Center, Royan Institute for Reproductive Biomedicine, ACECR, Tehran, Iran
3- Department of Andrology, Reproductive Biomedicine Research Center, Royan Institute for Reproductive Biomedicine, ACECR, Tehran, Iran

* Corresponding Author:
Anahita Mohseni Meybodi,
Reproductive Biomedicine
Research Center, Royan
Institute for Reproductive
Biomedicine, ACECR,
Tehran, Iran
E-mail:
Anahita.MohseniMeybodi
@lhsc.on.ca

Abstract

Background: Chlamydia trachomatis (CT) is one of the most prevalent sexually transmitted infections, causing genital tract infections and infertility. Defensins have an immunomodulatory function and play an important role in sperm maturation, motility, and fertilization. DEFB126 is present on ejaculated spermatozoa and is essential for them to pass through the female reproductive tract. The purpose of the study was to determine the frequency of the 2-nt deletion of the DEFB126 (rs11467417) in Iranian infertile males with a recurrent history of CT.

Methods: Semen samples of 1080 subfertile males were investigated. Among patients who had CT-positive results, sperm DNA from 50 symptomatic and 50 asymptomatic patients were collected for the DEFB126 genotype analysis. Additionally, a control group comprising 100 DNA samples from individuals with normal spermogram and testing negative for CT was included in the study. The PCR-sequencing for detecting the 2-nt deletion of the second exon of the DEFB126 was performed.

Results: The Chi-squared test comparing all three groups revealed no significant difference across the different genotypes. Moreover, no significant difference between the symptomatic and asymptomatic groups was seen. However, analysis within CT-positive patients and controls demonstrated significant difference between the frequencies of homozygous del/del.

Conclusion: The higher frequency of the 2-nt deletion of the DEFB126 in CT- positive patients suggests that the occurrence of mutations in the DEFB-126 may cause the impairment of the antimicrobial activity of the DEFB126 protein and consequently makes individuals more susceptible to infections such as CT.

Keywords: Chlamydia infection, Defensin gene, Infertility.
To cite this article: Haratian K, Borjian Boroujeni P, Sabbaghian M, Maghareh Abed E, Moazenchi M, Mohseni Meybodi A. DEFB126 2-nt Deletion (rs11467417) as a Potential Risk Factor for Chlamydia Trachomatis Infection and Subsequent Infertility in Iranian Men. J Reprod Infertil. 2024;25(1):20-27. https://doi.org/10.18502/jri.v25i1.15195.

Introduction

Reproductive tract infections play a significant role in the development of male and female infertility. Among male infertility factors, 13–15% are related to infections (1). Chlamydia trachomatis (CT) is one of the most prevalent sexually transmitted infections worldwide and a common cause of pathology in both men and women, causing several genital tract infections, ectopic pregnancies, and infertility (2, 3). Defensins are cationic antibacterial and antiviral peptides classified as α-defensin, β-defensins, and h-defensins. Humans have only α-defensins and β-

defensins. In addition to their direct antimicrobial functions, defensins have immunomodulatory functions and are involved in many physiological processes (4). Studies have shown that defensins are widely distributed in both the female reproductive tract (FRT) and male reproductive tract (MRT), playing a dual role in host defense and fertility protection. Moreover, a high expression level of β-defensins is found in MRT (4-6). It is highly expressed in the epididymal caput and corpus, indicating that defensins play an important role in sperm maturation. The expression of defensins in MRT varies with androgen levels, age (7-9), and the status of microbial invasion (10-12). They protect the male reproductive system from bacterial infections by neutralizing lipopolysaccharide (LPS) (13) and downregulating proinflammatory cytokines (14). In addition to sperm maturation, animal and clinical studies have shown that defensins play an important role in sperm motility and fertilization (15). As a broad-spectrum antimicrobial peptide without drug resistance, defensin has great potential for the development of new natural antimicrobial treatments for reproductive tract infections (4). However, increasing evidence has shown that defensins can not only inhibit microbial invasion but also promote the invasion and adhesion of some microorganisms in certain biological environments (16).

DEFB126 is a secretory glycoprotein, rich in cysteine, that is secreted by the principal epididymal epithelial cells and covers the entire surface of spermatozoa (5, 17). DEFB126 is present in ejaculated spermatozoa and is essential for them to pass through the female reproductive tract. This ability is related to terminal sialic residues that have negative charges, as the removal of these residues from DEFB126 results in a decrease in the number of spermatozoa that can successfully transit through the cervical mucus (18, 19). The DEFB126 glycoprotein plays a multifaceted role in various reproductive events. It is involved in crucial functions such as the storage of sperm in the tail of the epididymis, facilitating sperm movement through cervical mucus, protecting spermatozoa from the female immune system, as well as shielding them from enzymatic and microbial attacks. Additionally, it contributes to sperm capacitation, binds to the oviduct epithelium, and aids in the formation of a reservoir of spermatozoa within the oviduct (20, 21). Upon capacitation, DEFB126 is released from the sperm surface, which appears to be important for facilitating binding to the zona pellucida (5, 19). Sperm cells that are not covered by DEFB126 show reduced ability to reach the egg, which consequently leads to reduced fertility (6, 21). Human DEFB126 is located on the subtelomeric region of chromosome 20p13 and does not contain copy number variations (22). Several studies have de-monstrated that a two-nucleotide deletion in DEFB126 gene on both chromosomes (del/del) results in a frameshift mutation and produces a non-stop mRNA that is prone to degradation due to nonsense-mediated mRNA decay (6, 23, 24). Protein expression in males with the del/del genotype was significantly lower compared to the other genotypes.

DEFB126 is secreted by the principal epididymal epithelial cells and covers the entire surface of spermatozoa (5, 17). In addition to facilitating sperm movement in cervical mucus and capacitation, it is also involved in the protection of spermatozoa against the female immune system as well as against enzymatic and microbial attacks (20, 21). Several studies have demonstrated that homozygous 2-nt deletion of the DEFB126 gene (rs11467417) impairs DEFB126 protein synthesis (6, 25, 26). Protein expression in males with del/del genotype exhibited significantly lower levels compared to individuals with other genotypes (25). Considering the above-mentioned facts, the purpose of the current study was to investigate whether there is any association between the 2-nt deletion of the DEFB126 gene and CT infection in infertile Iranian males referred to Royan Institute.

Methods

Semen collection and Chlamydia trachomatis (CT) infection analysis: In this pilot case-control study, semen samples of 1080 subfertile males (aged 26–55 years) from the Infertility Clinic of Royan Institute were investigated between 2015 to 2017. Initially, the participants were divided into two primary groups based on their semen analysis results:

A) the control group consisting of patients with normal spermogram, and (B) the case group comprising patients with abnormal spermogram. The case group samples were checked for white blood cell (WBC) count and sperm parameters such as sperm count, motility, and morphology.

Patients with a history of antibiotic consumption in the last two months, positive for HIV, HBsAg, HCV Ab, varicocele, azoospermia, and history of epididymitis or epididymoorchitis were excluded

from this study.

Semen samples were collected by masturbation after three days of abstinence. Sperm concentration, motility, and morphology were assessed according to World Health Organization guidelines (2). Serological testing for CT and molecular methods for detecting ctDNA in semen samples have been described in detail in our previously published article (27). Samples that were positive by molecular and serological testing were included as CT-positive samples.

The individuals in the case group who were CT-positive and had an abnormal spermogram with high WBC (>1000 *per/ml*), low sperm count, low normal morphology, and low sperm motility were considered as symptomatic positive group. The portion of the main control group that exhibited no particular semen abnormalities but tested positive for CT was categorized as the third group, referred to as the infected asymptomatic group. The study was approved by the Ethical Review Board of the Royan Institute, following the principles outlined in the Helsinki Declaration (reference number: IR.ACECR.ROYAN.REC.1394.27). After obtaining informed consent from all patients enrolled, semen samples were collected for the study.

DEFB126 genotyping: In this pilot case-control study, among patients who had CT-positive results, sperm DNA from 50 symptomatic and 50 asymptomatic patients were collected for DEFB126 genotype analysis. Additionally, a control group comprising 100 DNA samples from individuals with normal spermogram and testing negative for CT was included in the study. Specific primers which were used to detect ctDNA as well as the primers amplifying the region of 2-nt deletion of the DEFB126 gene are shown in table 1.

Table 1. DEFB126 primers of the study

Primer name	Primer sequence (5'-3')	Product size
CT1-F	GGACAAATCGTATCTCGG	320
CT1-R	GAAACCAACTCTACGCTG	320
CT2-F	ATCCATTGCGTAGATCTCCG	517
CT2-R	GCCATGTCTATAGCTAAAGC	517
DEF-F	AAGAATGGTTGGGCAATGTGC	256
DEF-R	CCACCATGCTTTAATGAGTCGGG	256

The PCR-sequencing technique was performed to detect the 2-nt deletion of the second exon of the DEFB126 gene [NM_030931.4:c.317_318del, NP_112193.1:p.Pro106fs] according to the detailed protocol described previously by Boroujeni et al. (25). Briefly, after performing the PCR with specific primers on sperm DNA, PCR products were sent to Fazabiotech Company (Tehran, Iran) for being sequenced. The sequencing analysis was conducted in accordance with the Sanger sequencing standards, utilizing the ABI 3730XL Capillary Sequencer (ThermoFisher, US). The obtained results were then compared to the wild-type DEFB-126 gene sequence sourced from NCBI (https://www.ncbi.nlm.nih.gov). The extracted data were analyzed with BLAST (https://blast.ncbi.nlm.nih.gov/Blast.cgi) and Finch TV (https://digitalworld-biology.com/FinchTV). The sequences corresponding to the second exon of the DEFB126 gene in all of the CT-positive (case group) and the CT-negative (control group) infertile males were compared with the reference sequence (NC_000020.10) available on the NCBI-Gene site.

The wt/wt genotype refers to patients who do not have any deletions and exhibit a normal homozygous genotype (Figure 1A). In the wt/del geno-
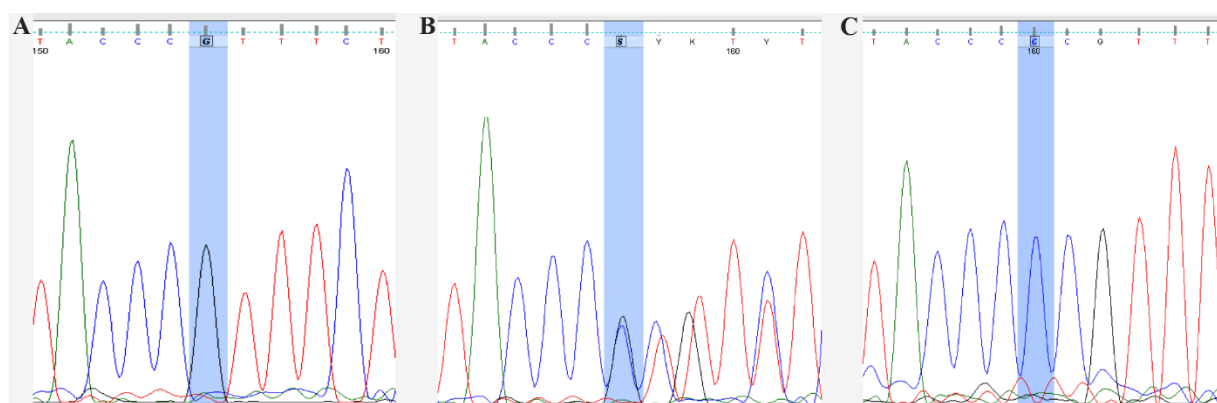
Figure 1. Sequence analysis chromatogram of wild-type (A), heterozygous (B), and del/del homozygous alleles (C) of DEFB126

type, the Finch TV analysis revealed the presence of two peak series in the corresponding regions. One peak represented the normal allele, while the other peak represented the mutant allele with a deletion of two cytosines, indicating a heterozygous genotype (Figure 1B). Patients with the del/del genotype demonstrated 2-nt deletion in both alleles in exon 2, indicating a homozygous genotype (Figure 1C).

Statistical analysis: Statistical analyses were carried out using SPSS version 22.0 (SPSS Inc., USA). Chi-squared test was used to determine the significance of the difference between DEFB126 mutation carriers and CT infection. A P-value of ≤ 0.05 was considered statistically significant.

Results

To determine the association between the DEFB-126 gene variant and CT infection, the presence of two cytosine deletions in exon 2 of the DEFB126 gene was evaluated in 100 CT-positive and 100 CT-negative infertile men.

Out of the 100 CT-positive males, 36% had a homozygous genotype for the wild-type allele, 37% had a heterozygous genotype with the deletion, and 27% had a homozygous genotype with the deletion. Among the control group, 41% exhibited a homozygous wild-type genotype, 48% had a heterozygous genotype, and 11% had a homozygous genotype with the deletion (Table 2). The Chi-squared test comparing all three groups revealed no significant differences across the different genotypes (p=0.074). Moreover, based on the statistical data, no significant difference between the two CT-positive groups (symptomatic *vs.* asymptomatic) was seen (p=0.307).

However, statistical analysis of CT-positive patients and controls demonstrated significant differences in the frequencies of homozygous del/del genotypes (p=0.042) (Table 2).

Based on the results of DEFB126 protein analysis (25), it was previously revealed that heterozygous wt/del individuals retain DEFB126 protein on their sperm surface. Therefore, by summing the frequencies of individuals with at least one normal allele (wt/del and wt/wt individuals), a significant difference was observed between the number of individuals with a wild-type allele (73%) and those without a copy of the wild-type allele (homozygous for the deletion, 27%) in infertile versus fertile men (p=0.013) (Table 2). As shown in table 2 and based on the Chi-squared test results, homozygote deletion was seen in 26% and 28% of symptomatic and asymptomatic CT-positive patients, respectively; which were significantly higher than that observed in the controls (11%) (p=0.046).

Discussion

Evaluation of CT infection in men is important because they can remain asymptomatic and transfer the infection to their female partners (28). This infection can persist for up to several years within couples and affect their fertility (29). Accordingly, CT infection may be one of the reasons for male infertility in those patients with idiopathic infertility (30). Previous studies demonstrated that a decrease in sperm count, sperm motility and progression, as well as an increase in the number of dead sperm and teratozoospermia index were all significantly associated with the presence of CT infection in infertile men (31, 32). Moreover,

Table 2. Genotype distribution between CT-positive (symptomatic and asymptomatic) patients and controls

		CT-positive patients 100	CT-positive patients 100				Controls 100	p-value	
			Symptomatic 50		Asymptomatic 50				
Genotype									
At least one normal allele	No deletion	73 (73%)*	37 (74%)**	15 (30%)	36 (72%)**	21 (42%)	89 (89%)*,**	41 (41%)	*p=0.021 **p=0.046
	Hetero			22 (44%)		15 (30%)		48 (48%)	
No wild-type allele	Homo	27 (27%)*,**	13 (26%)		14 (28%)		11 (11%)*,**		*p=0.021 **p=0.046

* Significant difference when comparing CT-positive patients and controls in terms of the presence of at least one normal allele (p≤0.05)
** Significant differences when comparing all three groups of samples with respect to the presence of at least one normal allele (p≤0.05)

Vigil et al. suggested that another possible effect of CT on male fertility might be due to its transmission to female partners, and consequently, the inflammatory processes that promote the generation of anti-sperm antibodies in the female reproductive tract (33).

Defensins, which are cationic proteins, are primarily expressed by neutrophils and epithelial cells. They play a significant role in the innate immune response at mucosal surfaces, acting as crucial components in combating infectious pathogens (34). The protective role of β-defensins against CT infections in FRT has been widely discussed (35-40). However, there is only one study on the role of defensins in preventing CT infections in MRT (41). To the best of our knowledge, this publication does not specifically focus on β-defensins.

The results of this study showed a significant difference between the frequencies of homozygous del/del genotype in CT-positive patients and controls. Interestingly, when there was only one normal allele (heterozygote), CT-positive patients and controls did not show a significant difference. Conversely, the frequency of individuals homozygous for the deletion in both symptomatic and asymptomatic CT-positive patients was higher when compared to the control group. Based on our results, the antimicrobial role of DEFB126 was suggested for the first time. Since it was previously discovered that the sperm surface of heterozygous wt/del individuals is still covered by the DEFB126 protein (25), it can be concluded that DEFB126 may play an important role in the effectiveness of immune response against CT infection. These results are in agreement with those of the previous studies. According to Yenugu et al., DEFB118, which is similar to DEFB126 and has the characteristics of other β-defensins, contributes to epididymal innate immunity and protects the sperm against attack by microorganisms in the male and female reproductive tracts (8). Moreover, recombinant β-defensin 22, the rat homolog of DEFB126, has been previously characterized as an effective antimicrobial peptide against E. coli, S. aureus, and C. albicans (42, 43). It is suggested that the pathogenicity of gram-negative bacteria, such as E. coli and CT, is usually associated with the lipopolysaccharide (LPS) layer, which is the main component of their cell walls. In humans, LPS can provoke an innate immune response (44) and induce defensin expression in the reproductive tract. On the other hand, defensins can regulate the immune response of macrophages against gram-negative bacterial infection by neutralizing LPS (13). In cases where defensin expression is absent, such as in individuals with a homozygous del/del genotype in DEFB-126, this process of immune defense may be compromised.

This study did not show any differences between symptomatic and asymptomatic CT-positive patients regarding the DEFB126 genotypes. In addition, 11% of the CT-negative infertile patients with normal spermogram showed homozygosity for this deletion (del/del). Based on the available information, it can be inferred that the 2-nt deletion had no detrimental impact on the parameters considered as "symptoms" in the patients' spermogram, including sperm count, motility, and morphology. Although He et al. in 2022 suggested an association between this variant and unstable defensin protein on the sperm surface and idiopathic asthenozoospermia (45), Tollner et al. considered this variant as a common mutation that causes impaired sperm "function" and subfertility in idiopathic infertile men with normal spermogram. It has been observed that sperm from individuals with the del/del genotype appears normal when assessed using conventional measures of male fertility potential, including sperm count, motility, and the percentage of sperm with normal morphology (5, 6). These data are also in agreement with our previous study on the role of this variant in idiopathic male infertility (25). However, based on the results of Aram et al., the proportion of DEFB126-positive sperm was positively correlated with sperm motility and normal morphology. Based on their findings, full-length DEFB126 can increase sperm motility *in vitro*. However, no genotype analysis was performed in this study (46).

Conclusion

The higher prevalence of the 2-nt deletion in the DEFB126 gene among CT-positive patients suggests a potential association between mutations in the DEFB126 gene and a compromised antimicrobial function of the DEFB126 protein. This compromise in the antimicrobial function of the DEFB126 protein may increase susceptibility to infections such as Chlamydia trachomatis (CT). Our results suggest that DEFB126 plays a significant role in the innate immune response against infectious pathogens. Like other genetic factors influencing individuals' immune defense mecha-

nisms, DEFB126 emerges as a noteworthy contributor to the modulation of Chlamydial infection pathogenesis and its potential impact on male fertility.

Conflict of Interest

The authors have no potential conflict of interest to declare.

References

1. Aitken RJ. The changing tide of human fertility. Hum Reprod. 2022;37(4):629-38.

2. World Health Organization. Infections, pregnancies, and infertility: perspectives on prevention. World Health Organization. Fertil Steril. 1987;47(6):964-8.

3. Brunham RC, Rekart ML. Considerations on Chlamydia trachomatis disease expression. FEMS Immunol Med Microbiol. 2009;55(2):162-6.

4. Zhai YJ, Feng Y, Ma X, Ma F. Defensins: defenders of human reproductive health. Hum Reprod Update. 2023;29(1):126-54.

5. Tollner TL, Bevins CL, Cherr GN. Multifunctional glycoprotein DEFB126—a curious story of defensinclad spermatozoa. Nat Rev Urol. 2012;9(7): 365-75.

6. Tollner TL, Venners SA, Hollox EJ, Yudin AI, Liu X, Tang G, et al. A common mutation in the defensin DEFB126 causes impaired sperm function and subfertility. Sci Transl Med. 2011;3(92):92ra 65.

7. Patrão MTCC, Silva EJR, Avellar MCWA. Androgens and the male reproductive tract: an overview of classical roles and current perspectives. Arq Bras Endocrinol Metabol. 2009;53(8):934-45.

8. Yenugu S, Hamil KG, Radhakrishnan Y, French FS, Hall SHJE. The androgen-regulated epididymal sperm-binding protein, human β-defensin 118 (DEFB118)(formerly ESC42), is an antimicrobial β-defensin. Endocrinology. 2004;145(7):3165-73.

9. Zhao Y, Diao H, Ni Z, Hu S, Yu H, Zhang Y, et al. The epididymis-specific antimicrobial peptide β-defensin 15 is required for sperm motility and male

fertility in the rat (Rattus norvegicus). Cell Mol Life Sci. 2011;68(4):697-708.

10. Fleming DC, King AE, Williams AR, Critchley HOD, Kelly RW. Hormonal contraception can suppress natural antimicrobial gene transcription in human endometrium. Fertil Steril. 2003;79(4):856-63.

11. Cao D, Li Y, Yang R, Wang Y, Zhou Y, Diao H, et al. Lipopolysaccharide-induced epididymitis disrupts epididymal beta-defensin expression and inhibits sperm motility in rats. Biol Reprod. 2010;83 (6):1064-70.

12. Quayle AJ, Porter EM, Nussbaum AA, Wang YM, Brabec C, Yip KP, et al. Gene expression, immunolocalization, and secretion of human defensin-5 in human female reproductive tract. Am J Pathol. 1998;152(5):1247-58.

13. Wilson SS, Wiens ME, Smith JG. Antiviral mechanisms of human defensins. J Mol Biol. 2013;425 (24):4965-80.

14. Pivarcsi A, Nagy I, Koreck A, Kis K, Kenderessy-Szabo A, Szell M, et al. Microbial compounds induce the expression of pro-inflammatory cytokines, chemokines and human β-defensin-2 in vaginal epithelial cells. Microbes Infect. 2005;7(9-10):1117-27.

15. Diao R, Fok KL, Chen H, Yu MK, Duan Y, Chung CM, et al. Deficient human β-defensin 1 underlies male infertility associated with poor sperm motility and genital tract infection. Sci Transl Med. 2014;6 (249):249ra108.

16. Xu D, Lu W. Defensins: a double-edged sword in host immunity. Front Immunol. 2020;11:764.

17. Yamaguchi Y, Nagase T, Makita R, Fukuhara S, Tomita T, Tominaga T, et al. Identification of multiple novel epididymis-specific β-defensin isoforms in humans and mice. J Immunol. 2002;169 (5):2516-23.

18. Gilks CB, Reid PE, Clement PB, Owen DA. Histochemical changes in cervical mucus-secreting epithelium during the normal menstrual cycle. Fertil Steril. 1989;51(2):286-91.

19. Tollner TL, Yudin AI, Tarantal AF, Treece CA, Overstreet JW, Cherr GN. Beta-defensin 126 on the surface of macaque sperm mediates attachment of sperm to oviductal epithelia. Biol Reprod. 2008; 78(3):400-12.

20. Toshimori K, Araki S, Öra C, Eddy EJ. Loss of sperm surface sialic acid induces phagocytosis: an assay with a monoclonal antibody T21, which recognizes a 54K sialoglycoprotein. Arch Androl. 1991;27(2):79-86.

21. Yudin AI, Treece CA, Tollner TL, Overstreet JW, Cherr GN. The carbohydrate structure of DEFB-

126, the major component of the cynomolgus Macaque sperm plasma membrane glycocalyx. J Membr Biol. 2005;207(3):119-29.

22. Hollox EJ, Barber JC, Brookes AJ, Armour JAJ. Defensins and the dynamic genome: what we can learn from structural variation at human chromosome band 8p23. Genome Res. 2008;18(11):1686-97.

23. Frischmeyer PA, Van Hoof A, O'Donnell K, Guerrerio AL, Parker R, Dietz HCJS. An mRNA surveillance mechanism that eliminates transcripts lacking termination codons. Science. 2002;295 (5563):2258-61.

24. Van Hoof A, Frischmeyer PA, Dietz HC, Parker R. Exosome-mediated recognition and degradation of mRNAs lacking a termination codon. Science. 2002;295(5563):2262-4.

25. Boroujeni PB, Ebrahimian S, Abedini M, Chayjan MR, Hassani M, Gourabi H, et al. The role of DEFB126 variation in male infertility and medically assisted reproduction technique outcome. Reprod Biomed Online. 2019;39(4):649-57.

26. Xin A, Cheng L, Diao H, Wu Y, Zhou S, Shi C, et al. Lectin binding of human sperm associates with DEFB126 mutation and serves as a potential biomarker for subfertility. Sci Rep. 2016;6:20249.

27. Moazenchi M, Totonchi M, Salman Yazdi R, Hratian K, Mohseni Meybodi M, Ahmadi Panah M, et al. The impact of Chlamydia trachomatis infection on sperm parameters and male fertility: A comprehensive study. Int J STD AIDS. 2018;29 (5):466-73.

28. Wollenhaupt J, Kolbus F, Weissbrodt H, Schneider C, Krech T, Zeidler HJC, et al. Manifestations of Chlamydia induced arthritis in patients with silent versus symptomatic urogenital chlamydial infection. Clin Exp Rheumatol. 1995;13(4):453-8.

29. Cunningham KA, Beagley KW. Male genital tract chlamydial infection: implications for pathology and infertility. Biol Reprod. 2008;79(2):180-9.

30. Matzuk MM, Lamb DJ. Genetic dissection of mammalian fertility pathways. Nat Med. 2002;4 Suppl:s41-9.

31. Idahl A, Abramsson L, Kumlin U, Liljeqvist J, Olofsson JI. Male serum Chlamydia trachomatis IgA and IgG, but not heat shock protein 60 IgG, correlates with negatively affected semen characteristics and lower pregnancy rates in the infertile couple. Int J Androl. 2007;30(2):99-107.

32. Veznik Z, Pospisil L, Svecova D, Zajicova A, Unzeitig V. Chlamydiae in the ejaculate: their influence on the quality and morphology of sperm. Acta Obstet Gynecol Scand. 2004;83(7):656-60.

33. Vigil P, Morales P, Tapia A, Riquelme R, Salgado AM. Chlamydia trachomatis infection in male partners of infertile couples: incidence and sperm function. Andrologia. 2002;34(3):155-61.

34. Lehrer RI, Lu W. α-Defensins in human innate immunity. Immunol Rev. 2012;245(1):84-112.

35. Wiechuła B, Cholewa K, Ekiel A, Romanik M, Doleżych H, Martirosian G. [HBD-1 and hBD-2 are expressed in cervico-vaginal lavage in female genital tract due to microbial infections]. Ginekol Pol. 2010;81(4):268-71. Polish.

36. Wiechuła BE, Friedek DA, Ekiel AM, Romanik MK, Martirosian G. Human neutrophil peptides in vaginitis/cervicitis of different etiology. Pol J Microbiol. 2007;56(3):185-9.

37. Wiesenfeld HC, Heine RP, Krohn MA, Hillier SL, Amortegui AA, Nicolazzo M, et al. Association between elevated neutrophil defensin levels and endometritis. J Infect Dis. 2002;186(6):792-7.

38. Mukura LR, Hickey DK, Rodriguez-Garcia M, Fahey JV, Wira CR. Chlamydia trachomatis regulates innate immune barrier integrity and mediates cytokine and antimicrobial responses in human uterine ECC-1 epithelial cells. Am J Reprod Immunol. 2017;78(6).

39. Noda-Nicolau NM, Bastos LB, Bolpetti AN, Pinto GVS, Marcolino LD, Marconi C, et al. Cervicovaginal levels of human β-defensin 1, 2, 3, and 4 of reproductive-aged women with Chlamydia trachomatis infection. J Low Genit Tract Dis. 2017;21(3): 189-92.

40. Fichorova RN, Morrison CS, Chen PL, Yamamoto HS, Govender Y, Junaid D, et al. Aberrant cervical innate immunity predicts onset of dysbiosis and sexually transmitted infections in women of reproductive age. PLoS One. 2020;15(1):e0224359.

41. Porter E, Yang H, Yavagal S, Preza GC, Murillo O, Lima H, et al. Distinct defensin profiles in Neisseria gonorrhoeae and Chlamydia trachomatis urethritis reveal novel epithelial cell-neutrophil interactions. Infect Immun. 2005;73(8):4823-33.

42. Liu H, Yu H, Gu Y, Xin A, Zhang Y, Diao H, et al. Human beta-defensin DEFB126 is capable of inhibiting LPS-mediated inflammation. Appl Microbiol Biotechnol. 2013;97(8):3395-408.

43. Diao H, Yu HG, Sun F, Zhang YL, Tanphaichitr N. Rat recombinant β-defensin 22 is a heparin-binding protein with antimicrobial activity. Asian J Androl. 2011;13(2):305-11.

44. Silva EJR, Ribeiro CM, Mirim AF, Silva AA, Romano RM, Hallak J, et al. Lipopolysaccharide and lipotheicoic acid differentially modulate epididymal cytokine and chemokine profiles and sperm

parameters in experimental acute epididymitis. Sci Rep. 2018;8(1):103.

45. He JY, Peng JY, Li QF, Lin XL, Cui YR, Ma SY, et al. DEFB126 polymorphisms and association with idiopathic asthenozoospermia in China. Asian J Androl. 2022;24(6):607-14.

46. Aram R, Chan PT, Cyr DG. Beta-defensin126 is correlated with sperm motility in fertile and infertile men. Biol Reprod. 2020;102(1):92-101.