

Effect of Omega-3 Supplementation on Visfatin, Adiponectin, and Anthropometric Indices in Women with Polycystic Ovarian Syndrome

Azadeh Nadjarzadeh ^{1,2}, Razieh Dehghani-Firouzabadi ³, Hoorieh Daneshbodi ^{4*}, Mohammad Hassan Lotfi ⁵, Niloofar Vaziri ⁴, Hassan Mozaffari-Khosravi ²

1- Nutrition and Food Security Research Centre, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

2- Department of Nutrition, Faculty of Health, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

3- Research and Clinical Center for Infertility, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

4- International Campus, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

5- Department of Biostatistics & Epidemiology, Faculty of health, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

Abstract

Background: Polycystic ovary syndrome (PCOS) is a multifactorial, metabolic disorder. Characteristics are chronic anovulation, polycystic ovaries and hyperandrogenism. The aim of this study was to determine the effect of omega-3 supplementation on visfatin, adiponectin, and anthropometric indices in PCOS women.

Methods: The study was a randomized double blind placebo-controlled clinical trial. It was conducted on 84 women with polycystic ovary syndrome (26.92 ± 5.05 years, $BMI = 31.69 \text{ Kg/m}^2$) who referred to the fertility and infertility research center and Shahid Sadoughi hospital in Yazd. After the examination, evaluation and para-medical assessment by obstetrician, they were recruited. They took 3 capsules of omega-3 (each one contained 180 mg EPA and 120 mg DHA) or placebo (each contained 1 g paraffin) daily for 8 weeks. Statistical analysis was paired T-test and student T-test, and a $p < 0.05$ was considered statistically significant.

Results: After the intervention, visfatin concentration did not change in neither groups. But, at the end of the study, the mean of adiponectin concentration increased ($p < 0.001$) in omega-3 group. Moreover, the mean of changes in this factor was significantly different between groups ($p < 0.005$). FSH did not change in two groups of the study. However, the mean of LH decreased about 1.74 mIU/ml in omega-3 group ($p < 0.005$). The mean of change of LH/FSH ratio between groups was significant ($p < 0.05$). After the intervention, prolactin did not meaningfully change in both groups.

Conclusion: Our results showed that 8 weeks of supplementation of omega-3 may have some beneficial effects on PCOS biochemical characteristics such as LH, LH/FSH, and adiponectin.

Keywords: Adiponectin, Fertility, FSH, LH, Obesity, Omega-3, PCOS, Visfatin.

To cite this article: Nadjarzadeh A, Dehghani-Firouzabadi R, Daneshbodi H, Lotfi MH, Vaziri N, Mozaffari-Khosravi H. Effect of Omega-3 Supplementation on Visfatin, Adiponectin, and Anthropometric Indices in Women with Polycystic Ovarian Syndrome J Reprod Infertil. 2015;16(4):212-220.

* Corresponding Author:
Hoorieh Daneshbodi,
International Campus of
Shahid Sadoughi
University of Medical
Sciences, Yazd, Iran
E-mail:
h_daneshbodi@ssu.ac.ir

Received: Nov. 26, 2014

Accepted: Mar. 3, 2015

Introduction

Polycystic ovary syndrome (PCOS) is a multifactorial, complex genetic, endocrine and metabolic disorder. Characteristics are chronic anovulation, polycystic ovaries and hyperandrogenism (1). According to different criteria, 4% to

8% of women exhibit PCOS in the world (2). The prevalence of PCOS in Iran was 14.6% using Rott's definition (3). PCOS has a tremendous negative impact on the physiology and metabolism of the body such as metabolic syndrome, in-

sulin resistance (IR), dyslipidemia, hypertension, hyperinsulinemia, abdominal obesity, type 2 diabetes mellitus (4), endometrial hyperplasia, cardiovascular disease (5), eating and mood disorders (6).

Omega-3 fatty acids are mainly found in plant oils and marine (7). It is beneficial for health and has protective effects on cardiovascular disease (8) and metabolic syndrome (9). In recent years, intake of omega-3 in diets is less than omega-6, and this imbalance in omega-6 to omega-3 ratio can be related to chronic diseases (10).

Symptoms of PCOS differ among women with PCOS and vary over time in each woman in the presence of particular precipitating factors, the most significant of which is an alteration in body weight (11, 12). There is evidence that obese PCOS women have more severe hyperandrogenism and a significantly higher incidence of anovulatory cycles, oligomenorrhea and/or hirsutism compared to normal weight ones (12) and weight has an effect on the levels of androgens (13), adiponectin, and visfatin (14). Belzung et al. demonstrated the highest dose of omega-3 causes the more reduction in fat mass (16). Campbell et al. showed that the impact of omega-3 on obesity is obvious (17).

Visfatin is an adipokine that mimics the effect of insulin and is secreted by visceral fat tissue (18) and it is related to insulin resistance (19). Visfatin is related to Body Mass Index (BMI) (20) and Waist to Hip Ratio (WHR) (21). Some studies showed that visfatin increased in subjects with abdominal obesity and PCOS (19, 22, 23). Visfatin similarity to insulin may stimulate the androgen synthesis and be the pathogenesis of PCOS (24). The regulatory effect of omega-3 on the level of some adipokines like leptin was evaluated (25), but its effects on visfatin levels was limited (26).

Adiponectin, as another adipokine, improves insulin sensitivity and has an anti-inflammatory effect (27). Some studies showed a decrease in adiponectin level in PCOS women (28). Some studies demonstrated that the low level of adiponectin can be related to pathogenesis of PCOS and its metabolic complications (29). Mohammadi et al. showed the effect of omega-3 on elevating the level of adiponectin in PCOS women (30).

Release of LH in PCOS women is more irregular than other women (31). Elevation in serum LH

and LH:FSH ratio in PCOS women is common (32, 33). Also, prolactin level is higher in these women despite FSH level (34). Some studies showed the effect of omega-3 on treatment of PCOS (35). Others showed no effect on LH and FSH after supplementation by polyunsaturated fatty acid (PUFA) (36, 37).

There were many studies about the effect of omega-3 on metabolic statuses of many diseases (38-40), but studies on PCOS patients were limited (41). Understanding the effects of omega-3 on obesity and some adipokines can be critical. The objective of this study was evaluating the effect of omega-3 supplementation on these hormone profiles, some adipokines, and some anthropometric indices in PCOS women.

Methods

The study was a randomized double blind placebo-controlled clinical trial and was executed from November 2011 to May 2012. It was approved by Shahid Sadoughi University of Medical Sciences' ethics committee (IRB no 50338), Yazd, Iran. This study was a part of the large trial registered in Iranian Registry of Clinical Trial (Registry no IRCT2012071410281N1). It was conducted on 84 women with polycystic ovary syndrome (26.92 ± 5.05 years, $BMI=31.69 \text{ Kg/m}^2$), referred to the fertility and infertility research center and Shahid Sadoughi hospital in Yazd. After the examination, evaluation and para-medical assessment by obstetrician, they were recruited. Inclusion criteria were: PCOS diagnostic criteria, age between 20-40 years, and BMI of 25-40. Exclusion criteria included metabolic disease, thyroid disease, hyperprolactinemia, hypercortisolemia, congenital adrenal hyperplasia, Cushing's syndrome, pituitary disorder, neoplasm, renal and liver diseases, history about intake of any drug during the last 3 months that may affect the insulin sensitivity or hormonal profile such as: oral contraceptives, glucocorticoids, ovulation induction agents, anti-diabetic and anti-obesity drugs, estrogenic, anti-androgenic or anti-hypertensive medication, being in any diet in the last 6 months, any addiction to tobacco and alcohol, taking omega-3 supplement in the last 3 months, menopause, allergy to sea foods, and the use of anti-coagulant medicine. And the participants were excluded if they needed OCP for treatment, were not willing to be in the study, did not obey the protocols of the study, or could not digest more than 20% of supplements.

The Rotterdam ESHRE/ASRM criteria for confirming the diagnosis of PCOS were: oligo-anovulation, clinical/or biochemical evidence of hyperandrogenism, and polycystic ovaries on ultrasound examination (11). Thorough clinical examination with special focus on clinical hyperandrogenism (*i.e.* hirsutism and or hypergonadism) was done.

After explaining the objectives of the study to the patients, they signed the consent form. The 24 hr recall was completed for each patient before and after the intervention to eliminate the effect of change in dietary intake. Recall's data were analyzed using Nutritionist 4 software (N-Squared Inc, San Bruno, CA, USA). Subjects were randomly divided in to two groups of A and B to reduce the bias. Researchers didn't know which groups take the omega-3 supplement or placebo until the end of the study. Both groups were given three capsules of omega-3 or placebos every day for 8 weeks (42). Each capsule of omega-3 contained 180 mg EPA and 120 mg DHA, and each placebo included 1 g paraffin (Zahravi, Iran). The appearance of placebo and omega-3 capsules were similar. All subjects were followed weekly by phone call, and they referred to the office bi-monthly to take the supplements to reduce the drop out. All participants were asked to maintain their usual diet and lifestyle habits.

For biochemical analysis, 10 ml of blood were collected from all samples before and after the intervention. Serum was kept in -80°C until analysis. Serum levels of visfatin, adiponectin, LH, FSH, and prolactin were measured. The LH/FSH ratio was calculated. Serum levels of visfatin (Cusabio, China), adiponectin (Medignost, Germany), LH (DiaMetra, Germany), FSH (DiaMetra, Germany), and prolactin (DiaMetra, Germany) were measured by ELISA Reader (Statfax2600, USA).

Weight, height, and BMI (weight in kg divided by height in m^2) were measured before the intervention for checking the inclusion criteria.

Weight (kg) was measured twice to the nearest 0.1 kg, with no shoes. Height (m) was measured twice to the nearest 0.5 cm, with no shoes.

Statistical analysis included mean and standard deviation, percentage, ratio, chi-square, pearson correlation coefficient test, paired T-test, student T-test, ANOVA, Wilcoxon, and Mann-whitney. SPSS 12 Software was used (SPSS Inc, Chicago IL) for analyzing the data and a p-value less than 0.05 was considered statistically significant.

Results

In this study, one of the participants became pregnant and left the study. Five participants dropped out because of taking less than 20% of our capsules. The remaining 78 participants completed the study. Finally, 78 people (92.8%) among the patients, 39 patients in the group receiving omega-3 and 39 patients in the group receiving placebo, finished the eight week trial.

Mean of age was 26.9 ± 5.9 years in omega-3 and 26.9 ± 5.0 years in placebo groups. Mean of BMI was $31.46 \pm 5.74 \text{ Kg/m}^2$ in omega-3 and $31.88 \pm 3.86 \text{ Kg/m}^2$ in placebo group. There were not any significant differences between groups in terms of age, weight, and other characters before the study.

There were no significant differences between and within groups in energy, macronutrients, omega-3, omega-6, and omega-6/mega-3 daily intake in the beginning of the study and after the intervention (Table 1).

Table 2 shows the mean of BMI and WHR in both groups. There were no significant differences between groups before the study. BMI remained unchanged in both groups at the end of the study. Significant increase in WHR was obtained in omega-3 group over the 8 weeks in comparison to baseline values ($p < 0.005$). WHR remained unchanged in placebo group after the intervention. Before the study, there was not a meaningful difference in terms of mean of visfatin, adiponectin, FSH, LH, prolactin concentration, and LH/FSH ratio in serum between groups. After the intervention, mean of visfatin increased up to 0.07 ng/ml in omega-3 group but the increasing level was not significant ($p = 0.34$). There was not a significant change ($p = 0.61$) in placebo group, too. Also, the mean of change between groups after the study was not significant ($p = 0.82$).

At the end of the study, mean of adiponectin significantly increased ($p < 0.001$) in supplement group. The mean of change of this factor was significantly different between groups ($p < 0.005$).

After 8 weeks of supplementation, no significant change in FSH concentration was shown in supplement and placebo groups ($p = 0.83$, $p = 0.87$, respectively). Also, the mean of change between groups after the study was not significant ($p = 0.79$).

The mean of LH decreased ($p < 0.005$) in omega-3 group, but there was not a significant change in placebo one ($p = 0.87$).

Table 1. Mean and standard deviation of intake of omega-3, omega-6, omega-6/omega-3, energy, and macronutrients between both groups before and after the intervention

		Placebo group N=39		Supplement group N=39		P-value (between groups)
		SD±Mean	Median	SD±Mean	Median	
Omega-3 (g/d)						
	Before the intervention	9.9±24.4	0.7	5.4±15.8	0.8	0.6 ***
	After the intervention	14.5±33.5	0.9	9.5±28.9	0.7	0.7 ***
	P-value *	0.34	--	0.6	--	--
Omega-6 (g/d)						
	Before the intervention	32.1±36.9	19.6	25.4±22.4	19.1	0.9 ***
	After the intervention	38.1±52.4	19.6	33.3±45.5	21.4	0.4 ***
	P-value *	0.4	--	0.1	--	--
Omega-6/omega-3						
	Before the intervention	26.0±20.4	--	22.9±11.4	--	0.4 ****
	After the intervention	23.4±20.0	--	25.1±12.4	--	0.6 ****
	P-value **	0.1	--	0.1	--	--
Energy (Kcal/d)						
	Before the intervention	2020.2±262.1	--	1970±235.3	--	0.4 ****
	After the intervention	1998.5±213.6	--	1960.6±223.8	--	0.5 ****
	P-value **	0.3	--	0.6	--	--
Fat (g/d)						
	Before the intervention	78.4±19.1	--	79.0±19.2	--	0.8 ****
	After the intervention	76.5±20.2	--	79.4±20.2	--	0.5 ****
	P-value **	0.5	--	0.5	--	--
Protein (g/d)						
	Before the intervention	63.5±11.6	--	59.8±10.5	--	0.1 ****
	After the intervention	65.1±12.8	--	59.6±9.3	--	0.3 ****
	P-value **	0.3	--	0.8	--	--
Carbohydrate						
	Before the intervention	268.6±58.6	--	260.2±32.3	--	0.5 ****
	After the intervention	266.5±41.2	--	259.2±29.1	--	0.3 ****
	P-value **	0.9	--	0.6	--	--

* Wilcoxon test, ** Paired sample t-test, *** Mann-Whitney U test, **** Student t test

The mean of prolactin did not meaningfully change in both groups (supplement group $p=0.62$, and placebo group $p<0.05$). After 8 weeks of the study, the mean of change of prolactin between groups was not significant ($p=0.08$) (Table 2).

The mean of LH/FSH ratio decreased ($p<0.001$) in supplement group, but there was not a significant change in placebo group ($p=0.98$). The mean of change of LH/FSH ratio between groups after the study was significant ($p<0.05$) (Table 2).

Discussion

The results of our study showed that omega-3 treatment was associated with significant improvement with LH, adiponectin concentration, WHR, and LH/FSH ratio in PCOS.

According to this study, no changes were seen in energy and macronutrient intakes and BMI in both groups after the intervention. These findings were similar to previous studies (30, 43). The study of Mohammadi et al. showed no changes in

Table 2. Comparison of quantitative variables between both groups before and after intervention

		Placebo group N=39		Supplement group N=39		P-value **
		SD±Mean	Median	SD±Mean	Median	
BMI (Kg/m²)						
	Before the intervention	31.88±3.86	--	31.46±5.74	--	0.75
	After the intervention	31.83±3.68	--	31.17±5.93	--	0.56
	P-value *	0.70	--	0.09	--	--
	Mean of change ***	-0.05±0.89	--	-0.29±1.04	--	0.28
WHR (cm)						
	Before the intervention	102.59±10.75	--	100.55±14.36	--	0.48
	After the intervention	102.27±10.20	--	98.77±14.55	--	0.22
	P-value *	0.53	--	<0.005	--	--
	Mean of change ***	-0.32±3.17	--	-1.78±3.21	--	0.28 *****
Visfatin (ng/ml)						
	Before the intervention	6.97±2.46	--	6.91±2.41	--	0.92
	After the intervention	7.01±2.60	--	6.98±2.51	--	0.96
	P-value *	0.61	--	0.34	--	--
	Mean of change ***	0.04±0.52	--	0.07±0.44	--	0.82
Adiponectin (ng/ml)						
	Before the intervention	5.16±3.93	--	4.44±1.92	--	0.31
	After the intervention	4.65±3.14	--	5.62±2.68	--	0.15
	P-value *	0.05	--	<0.005	--	--
	Mean of change ***	-0.51±1.60	--	1.17±2.10	--	<0.01
FSH (mIU/ml)						
	Before the intervention	5.60±2.22	--	5.75±3.07	--	0.80
	After the intervention	5.64±2.25	--	5.70±3.37	--	0.93
	P-value *	0.87	--	0.83	--	--
	Mean of change ***	-0.04±1.73	--	-0.05±1.50	--	0.79
LH (mIU/ml)						
	Before the intervention	6.60±3.62	--	8.18±3.87	--	0.07
	After the intervention	6.49±3.71	--	6.44±3.62	--	0.96
	P-value *	0.87	--	<0.01	--	--
	Mean of change ***	-0.11±4.28	--	-1.74±2.76	--	0.050
Prolactin (ng/ml)						
	Before the intervention	332.13±221.88	--	366.38±187.73	--	0.46
	After the intervention	388.08±197.97	--	353.18±178.02	--	0.42
	P-value *	0.05	--	0.62	--	--
	Mean of change ***	55.95±175.16	--	-13.20±165.14	--	0.08
LH/FSH ratio						
	Before the intervention	1.38±1.03	1.11	2.09±3.24	1.35	0.2
	After the intervention	1.33±0.85	1.33	1.33±0.92	1.05	0.8
	P-value ****	--	0.98	--	0.001	--
	Mean of change ***	-0.05±1.13	--	-0.76±2.94	--	0.02 *****

* Paired sample t-test, ** Independent sample t-test, *** Data after the study- first data, **** Wilcoxon test, ***** Mann-Whitney U test

WHR after 8 weeks of supplementation (30). However, some studies showed a reduction in WHR, BMI after 8 weeks of omega-3 supplementation in women with type 2 diabetes (26) and reduction of weight in mice (44, 45). Obesity and its comorbidities (diabetes, cancer, and heart disease) are linked to inflammation. Omega-3 had anti-inflammatory properties and has some effects on mediating receptors (46).

Our result about adiponectin level was compatible to previous studies in human (30, 47, 48) and animals (49, 50). The stimulation of Adipoq (adiponectin gene) in adipose tissue as a ligand of PPAR- γ (peroxisome proliferator-activated receptor γ) is one of the effects of EPA and DHA (44).

Evidence about the effect of omega-3 on visfatin level is limited. According to the study of Hajianfar et al., visfatin concentration increased after 8 weeks of the omega-3 supplementation (2 capsule/day, each capsule contain EPA 360 mg, DHA 240 mg) (26). In an *in vitro* study, it was suggested that AMPK activation has a role in up-regulation of visfatin gene after omega-3 intervention (51). This study showed no effect of omega-3 on visfatin level. The duration of the study and dose of EPA/DHA may not be enough to detect visfatin changes.

The improvement of LH concentration and LH/FSH ratio can cause the improvement of reproductive system. This trend can be explained by the mechanism related to arachidonic acids (52). Arachidonic acid can activate a steroidogenic acute regulatory protein (StAR). StAR is the rate-limiting step in steroidogenic pathway. High arachidonic acid can make 2-series prostaglandins (PGs), thromboxanes (TXs), and 4-series leukotrienes (LTs) and they induce StAR. However, high arachidonic acid can activate the LH-stimulated steroidogenesis. Increase of omega-3 in membrane can reduce the availability of arachidonic acid. So, StAR moves the cholesterol to the inner of mitochondrial membrane. It is the first and main step for steroidogenic pathway which results in production of androstenedione and testosterone (52).

Our data showed that omega-3 supplementation in 8 weeks has no effect on FSH concentration in PCOS women. These results are similar to the results of Kuzmanov et al.'s study. They examined 3 months of n-3 PUFA application on hormone status in 12 PCOS samples, but they did not see any changes on FSH concentration (53). Karakas et al.'s study showed similar results (36).

In our study, LH concentration and LH/FSH ratio significantly decreased after the intervention. Some studies showed no changes in LH concentration (52) and LH/FSH ratio after the intervention (37) like Karakas et al.'s study in using rich-PUFA diet for 3 months (36). Karakas suggested the intervention with more participants.

The concentration of prolactin after this study did not meaningfully change. No similar study about variation of prolactin in this case was found.

The strength of the current study was the clinical trial design. The limitation of our study was a shortness of intervention period and the clinical impacts were not collected. It is suggested that more clinical trials be conducted in this field with longer duration of intervention and in women with similar pattern of ovulations.

It has been suggested to design other studies with longer intervention. So, the effect of omega-3 could be more obvious. It has been suggested to measure the concentration of fatty acid on RBC membrane to check the protocol adherence and effect of omega-3 on other adipokines.

Conclusion

Omega-3 supplementation in PCOS women can show a benefit effect on adiponectin, LH concentration, WHR, and LH/FSH ratio. This effect was not associated with BMI, visfatin, FSH, and prolactin levels.

Acknowledgement

This study was derived from a master's dissertation. We thank the Research & Clinical Center of Infertility in Yazd for their support. We are extremely grateful to Maryam Chamari for her wise collaboration in this study.

Conflict of Interest

The authors declare that they have no conflict of interests.

References

1. Witchel SF. Puberty and polycystic ovary syndrome. *Mol Cell Endocrinol*. 2006;254-255:146-53.
2. Moran LJ, Hutchison SK, Norman RJ, Teede HJ. Lifestyle changes in women with polycystic ovary syndrome. *Cochrane Database Syst Rev*. 2011;(7): CD007506.
3. Tehrani FR, Simbar M, Tohidi M, Hosseini-panah F, Azizi F. The prevalence of polycystic ovary syndrome in a community sample of Iranian population: Iranian PCOS prevalence study. *Reprod Biol Endocrinol*. 2011;9:39.

4. Franks S, McCarthy MI, Hardy K. Development of polycystic ovary syndrome: involvement of genetic and environmental factors. *Int J Androl*. 2006;29(1): 278-85.
5. Xita N, Georgiou I, Tsatsoulis A. The genetic basis of polycystic ovary syndrome. *Eur J Endocrinol*. 2002;147(6):717-25.
6. Goodarzi MO, Dumesic DA, Chazenbalk G, Azziz R. Polycystic ovary syndrome: etiology, pathogenesis and diagnosis. *Nat Rev Endocrinol*. 2011;7(4): 219-31.
7. Kwintkiewicz J, Spaczynski RZ, Foyouzi N, Pehlivan T, Duleba AJ. Insulin and oxidative stress modulate proliferation of rat ovarian theca-interstitial cells through diverse signal transduction pathways. *Biol Reprod*. 2006;74(6):1034-40.
8. Park S, Park Y. Effects of dietary fish oil and trans fat on rat aorta histopathology and cardiovascular risk markers. *Nutr Res Pract*. 2009;3(2):102-7.
9. Poudyal H, Panchal SK, Diwan V, Brown L. Omega-3 fatty acids and metabolic syndrome: effects and emerging mechanisms of action. *Prog Lipid Res*. 2011;50(4):372-87.
10. Gallagher ML. Krause's food and nutrition therapy. 12th ed. St Louis, Missouri: Saunders Elsevier; 2008. Chapter 3, The nutrients and their metabolism; p. 39-143.
11. Allahbadia GN, Merchant R. Polycystic ovary syndrome and impact on health. *Middle East Fertil Soc J*. 2010;16(1):19-37.
12. Xita N, Tsatsoulis A. Review: Fetal programming of polycystic ovary syndrome by androgen excess: evidence from experimental, clinical, and genetic association studies. *J Clin Endocrinol Metab*. 2006; 91(5):1660-6.
13. Piouka A, Farmakiotis D, Katsikis I, Macut D, Gerou S, Panidis D. Anti-Mullerian hormone levels reflect severity of PCOS but are negatively influenced by obesity: relationship with increased luteinizing hormone levels. *Am J Physiol Endocrinol Metab*. 2009;296(2):E238-43.
14. Fantuzzi G. Adipose tissue, adipokines, and inflammation. *J Allergy Clin Immunol*. 2005;115(5): 911-9.
15. Hainault I, Carlotti M, Hajduch E, Guichard C, Lavau M. Fish oil in a high lard diet prevents obesity, hyperlipemia, and adipocyte insulin resistance in rats. *Ann N Y Acad Sci*. 1993;683:98-101.
16. Belzung F, Raclot T, Groscolas R. Fish oil n-3 fatty acids selectively limit the hypertrophy of abdominal fat depots in growing rats fed high-fat diets. *Am J Physiol*. 1993;264(6 Pt 2):R1111-8.
17. Sara CC, Bello NT. Omega-3 Fatty Acids and Obesity. *J Food Nutr Disord*. 2012;1:2.
18. Stofkova A. Resistin and visfatin: regulators of insulin sensitivity, inflammation and immunity. *Endocr Regul*. 2010;44(1):25-36.
19. Plati E, Kouskouni E, Malamitsi-Puchner A, Boutsikou M, Kaparos G, Baka S. Visfatin and leptin levels in women with polycystic ovaries undergoing ovarian stimulation. *Fertil Steril*. 2010;94(4): 1451-6.
20. Varma V, Yao-Borengasse A, Rasouli N, Bodles AM, Phanavanh B, Lee MJ, et al. Human visfatin expression: relationship to insulin sensitivity, intramyocellular lipids, and inflammation. *J Clin Endocrinol Metab*. 2007;92(2):666-72.
21. Chen MP, Chung FM, Chang DM, Tsai JC, Huang HF, Shin SJ, et al. Elevated plasma level of visfatin/pre-B cell colony-enhancing factor in patients with type 2 diabetes mellitus. *J Clin Endocrinol Metab*. 2006;91(1):295-9.
22. Ozkaya M, Cakal E, Ustun Y, Engin-Ustun Y. Effect of metformin on serum visfatin levels in patients with polycystic ovary syndrome. *Fertil Steril*. 2010;93(3):880-4.
23. Sandeep S, Velmurugan K, Deepa R, Mohan V. Serum visfatin in relation to visceral fat, obesity, and type 2 diabetes mellitus in Asian Indians. *Metabolism*. 2007;56(4):565-70.
24. Kowalska I, Straczkowski M, Nikolajuk A, Adamaska A, Karczewska-Kupczewska M, Otziomek E, et al. Serum visfatin in relation to insulin resistance and markers of hyperandrogenism in lean and obese women with polycystic ovary syndrome. *Hum Reprod*. 2007;22(7):1824-9.
25. Rossi AS, Lombardo YB, Lacorte JM, Chicco AG, Rouault Ch, Slama G, et al. Dietary fish oil positively regulates plasma leptin and adiponectin levels in sucrose-fed, insulin-resistant rats. *Am J Physiol Regul Integr Comp Physiol*. 2005;289(2): R486-R494.
26. Hajianfar H, Hosseinzadeh MJ, Bamonir A, Mohammad K, Askari GR, Entezari MH, et al. The effect of omega-3 on the serum visfatin concentration in patients with type II diabetes. *J Res Med Sci*. 2011;16(4):490-5.
27. Mamaghani F, Zarghami N, Malekiet MJ, Pourhasan-Moghaddam M, Hosseiniapanah F. Variation of adiponectin levels in normal and obese subjects: possible correlation with lipid profiles. *Int J Endocrinol Metab*. 2009;7(3):170-8.
28. Panidis D, Kourtis A, Farmakiotis D, Mouslech T, Rousso D, Koliakos G. Serum adiponectin levels in women with polycystic ovary syndrome. *Hum Reprod*. 2003;18(9):1790-6.
29. Escobar-Morreale HF, Villuendas G, Botella-Carretero JJ, Alvarez-Blasco F, Sanchon R, Luque-

- Ramirez M, et al. Adiponectin and resistin in PCOS: a clinical, biochemical and molecular genetic study. *Hum Reprod*. 2006;21(9):2257-65.
30. Mohammadi E, Rafrat M, Farzadi L, Asghari-Jafarabadi M, Sabour S. Effects of omega-3 fatty acids supplementation on serum adiponectin levels and some metabolic risk factors in women with polycystic ovary syndrome. *Asia Pac J Clin Nutr*. 2012;21(4):511-8.
 31. Barontini M, Garcia-Rudaz MC, Veldhuis JD. Mechanisms of hypothalamic-pituitary-gonadal disruption in polycystic ovarian syndrome. *Arch Med Res*. 2001;32(6):544-52.
 32. Spritzer PM, Poy M, Wiltgen D, Mylius LS, Capp E. Leptin concentrations in hirsute women with polycystic ovary syndrome or idiopathic hirsutism: influence on LH and relationship with hormonal, metabolic, and anthropometric measurements. *Hum Reprod*. 2001;16(7):1340-6.
 33. Banaszewska B, Spaczynski RZ, Pelesz M, Pawelczyk L. Incidence of elevated LH/FSH ratio in polycystic ovary syndrome women with normo- and hyperinsulinemia. *Rocz Akad Med Bialymst*. 2003;48:131-4.
 34. Wang Y, Yu P. [Clinical significance and changes of serum visfatin, adiponectin and leptin levels in patients with polycystic ovarian syndrome]. *Zhong Nan Da Xue Xue Bao Yi Xue Ban*. 2009;34(1):72-5. Chinese.
 35. Lydic M, Juturu V. Dietary approaches and alternative therapies for polycystic ovary syndrome. *Curr Nutr Food Sci*. 2008;4(4):265-81.
 36. Kasim-Karakas SE, Almario RU, Gregory L, Wong R, Todd H, Lasley BL. Metabolic and endocrine effects of a polyunsaturated fatty acid-rich diet in polycystic ovary syndrome. *J Clin Endocrinol Metab*. 2004;89(2):615-20.
 37. Kuzmanov AP. Do omega-3 fatty acids affect hormones related to polycystic ovary syndrome infertility? [master's thesis]. [United States]: University of Wyoming; 2010. 81 p.
 38. Ebbesson SO, Risica PM, Ebbesson LO, Kennish JM, Tejero ME. Omega-3 fatty acids improve glucose tolerance and components of the metabolic syndrome in Alaskan Eskimos: the Alaska Siberia project. *Int J Circumpolar Health*. 2005;64(4):396-408.
 39. Harris WS, Miller M, Tighe AP, Davidson MH, Schaefer EJ. Omega-3 fatty acids and coronary heart disease risk: clinical and mechanistic perspectives. *Atherosclerosis*. 2008;197(1):12-24.
 40. Anil E. The impact of EPA and DHA on blood lipids and lipoprotein metabolism: influence of apoE genotype. *Proc Nutr Soc*. 2007;66(1):60-8.
 41. Cussons AJ, Watts GF, Mori TA, Stuckey BG. Omega-3 fatty acid supplementation decreases liver fat content in polycystic ovary syndrome: a randomized controlled trial employing proton magnetic resonance spectroscopy. *J Clin Endocrinol Metab*. 2009;94(10):3842-8.
 42. Rafrat M, Mohammadi E, Asghari-Jafarabadi M, Farzadi L. Omega-3 fatty acids improve glucose metabolism without effects on obesity values and serum visfatin levels in women with polycystic ovary syndrome. *J Am Coll Nutr*. 2012;31(5):361-8.
 43. Vargas ML, Almario RU, Buchan W, Kim K, Karakas SE. Metabolic and endocrine effects of long-chain versus essential omega-3 polyunsaturated fatty acids in polycystic ovary syndrome. *Metabolism*. 2011;60(12):1711-8.
 44. Flachs P, Mohamed-Ali V, Horakova O, Rossmeisl M, Hosseinzadeh-Attar MJ, Hensler M, et al. Polyunsaturated fatty acids of marine origin induce adiponectin in mice fed a high-fat diet. *Diabetologia*. 2006;49(2):394-7.
 45. Nakatani T, Kim HJ, Kaburagi Y, Yasuda K, Ezaki O. A low fish oil inhibits SREBP-1 proteolytic cascade, while a high-fish-oil feeding decreases SREBP-1 mRNA in mice liver: relationship to anti-obesity. *J Lipid Res*. 2003;44(2):369-79.
 46. Oh DY, Talukdar S, Bae EJ, Imamura T, Morinaga H, Fan W, et al. GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects. *Cell*. 2010;142(5):687-98.
 47. Kondo K, Morino K, Nishio Y, Kondo M, Fuke T, Ugi S, et al. Effects of a fish-based diet on the serum adiponectin concentration in young, non-obese, healthy Japanese subjects. *J Atheroscler Thromb*. 2010;17(6):628-37.
 48. Itoh M, Suganami T, Satoh N, Tanimoto-Koyama K, Yuan X, Tanaka M, et al. Increased adiponectin secretion by highly purified eicosapentaenoic acid in rodent models of obesity and human obese subjects. *Arterioscler Thromb Vasc Biol*. 2007;27(9):1918-25.
 49. Duda MK, O'Shea KM, Lei B, Barrows BR, Azimzadeh AM, McElfresh TE, et al. Dietary supplementation with omega-3 PUFA increases adiponectin and attenuates ventricular remodeling and dysfunction with pressure overload. *Cardiovasc Res*. 2007;76(2):303-10.
 50. Higuchi T, Shirai N, Saito M, Suzuki H, Kagawa Y. Levels of plasma insulin, leptin and adiponectin, and activities of key enzymes in carbohydrate metabolism in skeletal muscle and liver in fasted ICR mice fed dietary n-3 polyunsaturated fatty acids. *J Nutr Biochem*. 2008;19(9):577-86.

51. Nettleton JA, Katz R. n-3 long-chain polyunsaturated fatty acids in type 2 diabetes: a review. *J Am Diet Assoc.* 2005;105(3):428-40.
52. Phelan N, O'Connor A, Kyaw Tun T, Correia N, Boran G, Roche HM, et al. Hormonal and metabolic effects of polyunsaturated fatty acids in young women with polycystic ovary syndrome: results from a cross-sectional analysis and a randomized, placebo-controlled, crossover trial. *Am J Clin Nutr.* 2011;93(3):652-62.
53. Kuzmanov A, Broughton KS. Role of marine omega-3 fatty acids in PCOS cancer risk. *Proceedings of the 13th World Congress on Controversies in Obstetrics, Gynecology & Infertility (COGI); 2010 November 4-7; Berlin, Germany: Monduzzi Editore; c2010. p. 349-351.*